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Int. J. Biol. Chem. Sci. 9(5): 2712-2718, October 2015

International Journal of Biological and Chemical Sciences

ISSN 1997-342X (Online), ISSN 1991-8631 (Print)

Original Paper

http://ajol.info/index.php/ijbcs http://indexmedicus.afro.who.int

Syntheses, characterizations and antioxidant activities of copper complexes of quercetin as influenced by redox states

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ABSTRACT

Copper is an essential element for living but imbalance between Cu^+ and Cu^{2+} produces free radicals. Quercetin is an effective metal chelator and its chelates are often better free radical scavengers than free quercetin. In this study, the syntheses, characterizations, antioxidant activities of quercetin as well as its Cu (I) and Cu (II) complexes using UV-VIS, IR spectrometric methods, and 2,2-diphenyl-1-picrylhydrazyl (DPPH) scavenging assay were investigated. 1:1 and 2:1 stoichiometric ratios were obtained for the dark-brown Cu⁺ and green Cu²⁺ quercetin complexes respectively. Bathochromic shifts from 268 nm (band II) and 362 nm (band I) in quercetin were observed at 302 nm and 386 nm for Cu (I) –quercetin complex and at 281 nm and 394 nm for Cu (II) - quercetin complex. FT-IR showed shifts in the positions of carbonyl and hydroxyl groups with presence of bands at 932.01 cm⁻¹ in Cu (I) – quercetin and 928.57 cm⁻¹ in Cu (II) – quercetin confirming complexation. Both complexes were better free radical scavengers than free quercetin with Cu (I)-quercetin complex are related to relative stability of fully filled orbital of Cu⁺ over Cu²⁺. © 2015 International Formulae Group. All rights reserved.

Keywords: Oxidation state, relative stability, DPPH, Stoichiometric ratio, complexation

INTRODUCTION

Copper (Cu) is a transition element that occurs as Cu^+ (cuprous) and Cu^{2+} (cupric). Cu^+ [3d¹⁰] is more stable than Cu^{2+} [3d⁹] because of stability of fully filled d-orbital (Olajire, 2005). Cu is a potent antimicrobial agent and is involved as a catalytic cofactor in hemoglobin formation, respiration, xenobiotic metabolism and antioxidant defense

mechanism (Mercer, 2001; Puig and Thiele, 2002; Omoto et al., 2005). As essential as Cu is to living organisms, it is also toxic because its ions are potent generators of free radicals (Puig and Thiele, 2002; Panhwar et al., 2010; Riha et al., 2014). These properties are driven by its ability to cycle between two redox states; oxidized Cu²⁺ and reduced Cu⁺ (Mercer, 2001; Omoto et al., 2005; Aaseth et

al., 2015). However, in the process of moving between Cu^+ and Cu^{2+} states, Cu imbalance is produced and leads to generation of free hydroxyl radicals which can damage lipids, proteins and DNA and disturb the synthesis of iron sulfur clusters (Festa and Thiele, 2011; Bukhari et al., 2008; Riha et al., 2014). Though each organism has in place mechanisms to safely excrete the excess Cuions in its body, this could be hampered when excess ions are produced. Therefore, chelation therapy can be used to remove excess Cu ions from vulnerable sites in critical organs (Omoto et al., 2005; Aaseth et al., 2015).

Chelation therapy is a promising method in treating pathological diseases arising from oxidative stress caused by excess or dysregulation of transition metals. This is achieved by suppressing the metal induced toxicity (Dehghan and Khoshkam, 2012; Riha et al., 2014). Chelating agents such as flavonoids have natural ways of complexing toxic metals and are easily excreted from the body. This plays a major role in limiting metal bioavailability and suppressing metal toxicity (Malešev and Kunti, 2007; Riha et al., 2014).

Flavonoids are known to possess biological and pharmacological properties such as anti-cancer, anti-ulcer, anti-allergic, antioxidant, antiviral and anti-inflammatory needed in cardiovascular protection (Bukhari et al., 2008; Olajire and Azeez, 2011). They are present in fruits, vegetables and have free radical scavenging ability (Malešev and Kunti, 2007; Panhwar et al., 2010; Dehghan and Khoshkam, 2012). They are effective metal chelators and their chelating ability is widely believed as another mechanism of their antioxidant activity. These chelates have been reported to possess higher potency against free radicals than free flavonoids as it has confirmed that the biological activity of organic ligands such as flavonoids can be increased when co-coordinated with suitable metal ion thereby providing better protection against oxidative stress. Flavonoids chelate using carbonyl oxygen and hydroxyl groups present in their molecules (Kostyuk et al.,

2004; Moridani et al., 2003; Malešev and Kunti, 2007; Pereira et al., 2007; Bukhari et al., 2008; Dehghan and Khoshkam, 2012).

Quercetin (3, 3', 4', 5, 7 pentahydroxyflavone) is one of the most common flavonols (a class in flavonoids) found in tea, onions, nuts, berries, cauliflower, and cabbage and it reacts with free radicals by donating its proton without itself becoming a radical. It is a strong metal chelator and a potent antioxidant compound possessing higher antioxidant activity than α -tocopherol, L-ascorbic acid and butylated hydroxylanisol (BHA) (Kostyuk et al., 2004; Moridani et al., 2003; Malešev and Kunti, 2007; Pereira et al., 2007; Olajire and Azeez, 2011; Dehghan and Khoshkam, 2012). It possesses three possible chelating sites in competition: the 3-hydroxy-4-carbonyl, the 5-hydroxy-4-carbonyl, and the 3', 4'-dihydroxyl (catechol) groups. Studies on complexation of metals using quercetin have been reported for Fe²⁺, Fe³⁺, Cu²⁺, Zn²⁺ and were found to be more effective against radicals than free quercetin. These complexes have shown better efficiencies in protecting red blood cells against asbestos-induced oxidative injury in vitro (Kostyuk et al., 2004; Moridani et al., 2003).

Since Cu is easily converted from Cu^{2+} to Cu^{1+} with its attendant toxicity, little or no study has reported the possible effects this conversion can have on the antioxidant activities of their chelates, hence, this study aimed at determining the effects of redox states on syntheses, spectra characterizations and antioxidant activities of copper complexes of quercetin.

MATERIALS AND METHODS Chemicals

2,2-diphenyl-1-picrylhydrazyl (DPPH), copper (II) sulphate pentahydrate (CuSO₄.5H₂O), copper (I) oxide (Cu₂O), methanol and ethanol. All reagents were analar grade and were bought from Sigma Aldrich, Germany. Deionized-distilled water was used all through the experiments.

Instrumentation

UV-visible spectra of quercetin, Cu (I) - quercetin and Cu (II) - quercetin complexes were obtained using Jenway 6405 UV-Visible spectrophotometer and Infrared spectra were recorded using KBr pellets in the spectral range 4000-400 cm⁻¹ on FT-IR Spectrophotometer (Model 500, Buck Scientific Inc.).

Stoichiometric ratios of copper ions with quercetin

Job's method of continual variation was used to determine the stoichiometric ratio between quercetin and the Cu ions. Equimolar concentration $(4x10^{-4}M)$ solutions of quercetin, Cu₂O and CuSO₄.5H₂O were prepared. Cu₂O and quercetin were mixed in different ratios varying from 1:9 to 9:1 and the absorbance was measured at 427 nm. The same procedure was used for CuSO₄.5H₂O and quercetin.

Synthesis

Cu (I) and Cu (II) complexes of quercetin were prepared using a modified method of Bukhari et al. (2008). For Cu₂O, solid quercetin (0.3 g, 1×10^{-3} mol) was dissolved in 20 ml methanol in a 50 ml round bottom flask placed on electromagnetic stirrer, this solution was mixed thoroughly until the quercetin completely dissolved to form yellow-colored solution, solid Cu₂O (0.143 g, 1×10^{-3} mol) was added quickly; the color of the solution turned dark brown with continued stirring for 2 hr at room temperature. After stirring, the solution was filtered and the filtrate was evaporated using rotary evaporator. A dark brown solid was obtained (0.41 g). Elemental analysis of the complex was C = 37.43%, H = 2.93% with molecular formula C15H14Cu2O10 and molecular mass 481.36 g.

For CuSO₄.5H₂O, solid quercetin (0.3 g, $1x10^{-3}$ mol) was dissolved in 20 ml methanol in a 50 ml round bottom flask placed on electromagnetic stirrer, this solution was mixed thoroughly until the quercetin

completely dissolved to form yellow-colored solution, solid $CuSO_4.5H_2O$ (0.355 g, $2x10^{-3}$ mol) was added quickly; the color of the solution turned green with continued stirring for 2 hr at room temperature. After stirring, solution was filtered and the filtrate was evaporated using rotary evaporator. Green solid (0.524 g) was obtained. Elemental analysis of the complex gave C = 30.05%, H = 4.15% with molecular formula $C_{17}H_{28}Cu_2O_{18}S$ and molecular mass 6799.55 g.

Determination of antiradical activity

The method of Olajire and Azeez (2011) was used for the determination of antioxidant activity. 1%, 0.8%, 0.6%, 0.4% and 0.2% solutions of quercetin, Cu (I) – quercetin and Cu (II) – quercetin complexes were prepared. 1 ml of each concentration was measured into test tubes and 4 ml of 0.004 M DPPH was added to each tube in a dark room. The blank was prepared by measuring 1 ml of methanol in test tube and 4 ml of DPPH was added. Both blank and samples were allowed to stand for 30 minutes in dark room before reading at wavelength 517 nm. Antioxidant activity was calculated using the equation below:

 $A_{(517)} = [(A_{blank} - A_{sample}) / A_{blank}] \times 100.$

RESULTS

Stoichiometric ratio and physical properties

Job's method of continual variation was used to determine the stoichiometric ratio as presented in Figure 1. Absorbance plots of Cu (I) - quercetin complex and Cu (II) – quercetin complex against mole fractions of quercetin have maximum absorptions at 0.4 for CuSO₄.5H₂O and at 0.52 for Cu₂O. These show a 2:1 stoichiometric ratio for Cu (II) quercetin and 1:1 stoichiometric ratio for Cu (I) - quercetin. Syntheses of the complexes were done using these ratios. Cu (I) quercetin and Cu (II) - quercetin complexes were dark brown and green solids respectively which were not soluble in water but soluble in ethanol and methanol.

UV-visible spectra characterization

Figure 2 shows the characteristic peaks of free quercetin, Cu (I) - quercetin and Cu (II) - quercetin complexes. In free quercetin, maximum absorptions (λ_{max}) were observed at 268 nm and 362 nm while Cu (I) -quercetin and Cu (II) - quercetin complexes have maximum absorptions (λ_{max}) at 302 nm and 386 nm; 281 nm and 394 nm respectively. In quercetin, band II with 268 nm is a characteristic (λ_{max}) absorption attributable to transition from π - π^* in the ring A consisting benzene ring while band I with 362 nm is characteristic λ_{max} of transition in the ring B consisting the cinnamoyl system and ranges from 300 – 400 nm. Shifts in λ_{max} from 268 nm in quercetin (band II) to 302 nm and 281 nm in Cu (I) - quercetin and Cu (II) quercetin complexes respectively were obtained. Also, Shifts in λ_{max} from 362 nm (band I) in free quercetin to 386 nm and 394 nm in Cu (I) - quercetin and Cu (II) quercetin complexes respectively were observed.

Infra-Red spectra characterization

The coordination sites and binding properties in free quercetin, Cu (I) - quercetin and Cu (II) – quercetin complexes are presented in Table 1. The stretching vibration of C=O in free quercetin occurred at 1705.09 cm⁻¹ which was shifted to 1718.33 cm⁻¹ and 1727.97 cm⁻¹ in Cu (I) - quercetin and Cu (II) quercetin complexes respectively. The shift in O-H was from 3418.37 cm⁻¹ in free quercetin to 3425.60 cm⁻¹ and 3453.53 cm⁻¹ in Cu (I) quercetin and Cu (II) - quercetin complexes respectively. The bands at 1388.91cm⁻¹, 1614.62 cm⁻¹ and 1226.57cm⁻¹ were related to v(C-OH), v(C=C) and v(C-O-C) in free quercetin which were shifted to 1373.56 cm⁻¹, 1612.31 cm⁻¹ and 1266.72 cm⁻¹ in Cu (I) quercetin complex and, 1363.18 cm⁻¹, 1616.73 cm⁻¹ and 1261.59 cm⁻¹ in and Cu (II) quercetin complex. The presence of υ (Cu-O) at 932.01 cm⁻¹ and 928.57 cm⁻¹ in Cu (I) quercetin and Cu (II) - quercetin complexes respectively was obtained.

Antioxidant activity

Figure 3 shows the antioxidant activities of free quercetin, Cu (I) - quercetin and Cu (II) - quercetin complexes. Cu (I) - quercetin complex had highest antioxidant activity (86.9%) followed by quercetin (76.4%) and Cu (II) - quercetin complex (73.9%). Inhibitory concentration at which 50% of the radicals were scavenged (IC₅₀) of quercetin, Cu (I) - quercetin and Cu (II) - quercetin complexes are 0.52 mg/ml, 0.39 mg/ml, 0.56 mg/ml respectively.



Figure 1: Absorbance plots of CuSO₄.5H₂O and Cu₂O against mole fractions of quercetin.



Figure 2: UV-Visible spectra of free quercetin Cu (I) - quercetin and Cu (II) - quercetin complexes.



Figure 3: Antioxidant activities of quercetin, Cu (I) - quercetin and Cu (II) - quercetin complexes.

Bands	Quercetin	Cu (I) - quercetin	Cu (II) - quercetin
υ (O-H)	3418.37 s	3425.60 s	3453.53 s
υ (C=O)	1705.09s	1718.33 s	1727.97 s
υ (C-H)	2931.31 s	2936.31 s	2996.84 s
υ (C-OH)	1388.91 m	1373.56 m	1363.18 m
υ (C-O)	1107.18 s	1112.22 s	1164.58 s
υ (C=C)	1614.62	1612.31 s	1616.73
υ (C-O-C)	1226.57 s	1266.72 m	1261.59 s
υ (Cu-O)		932.01 s	928.57
a Strong and m Madium			

Table 1: FT-IR bands (cm⁻¹) of quercetin, Cu (I) - quercetin and Cu (II) - quercetin complexes.

s - Strong and m - Medium.

DISCUSSION

Metal-flavonoid complexes are of high importance and are therefore characterized using different spectroscopic methods to understand their physical properties. Cu (I) quercetin and Cu (II) - quercetin complexes in this study were dark-brown and green colored respectively. This is supported by the evidence that metal-flavonoid complexes are often colored (Malešev and Kunti, 2007).

Formation of complexes was confirmed using UV-Vis and IR spectroscopic methods. Bathochromic shifts obtained in UV-Vis absorption bands of copper complexes in comparison with quercetin could have been caused by increased conjugative effects of Cu (I) and Cu (II) suggesting interactions between these metals and condensed rings of the flavonoids. It has been reported that bathochromic shifts are usually observed in the absorption bands of flavonoid in the presence of metal ions (Malešev and Kunti, 2007). This is in agreement with results reported by Bukhari et al. (2008) and Panwar et al. (2010) that indicate that these shifts are caused by the binding of the metal ion with flavonoid.

Also, shifts in υ (OH), and υ (CO) positions of quercetin and presence of v (Cu-O) which was absent in free quercetin indicate that the complexes were formed, and hydroxyl and carbonyl groups were involved in the complexation. It has been reported that hydroxyl and carbonyl groups at 3 and 4 positions are the first sites of chelation for flavonoid especially for flavonols while hydroxyl groups at 3' and 4' bind the second metal (Malešev and Kunti, 2007; Bukhari et al., 2008; Panhwar et al., 2010; Dehghan and Khoshkam, 2012). Therefore, in this study hydroxyl groups at 3, 3', 4'- positions and carbonyl group at 4 - positions were identified as the binding sites for Cu (II) - quercetin complex because it is 2:1 while 3-hydroxyl and 4-carbonyl positions were identified as the binding sites for Cu (I) - quercetin complex because it is 1:1.

Metal-flavonoid complexes have been reported to possess higher antioxidant against free radicals than free flavonoid. In this study, IC₅₀ of the complexes showed that they possess higher scavenging ability against free radicals than free quercetin with Cu (I) quercetin the most potent because the lower the IC_{50} the more potent the complex is (Olajire and Azeez, 2011). These results are in agreement with Kostyuk et al. (2004) who reported that flavonoid complexes are more potent free flavonoid in scavenging free radicals. Equally, increase in the antioxidative potencies of Cu (I) - quercetin and Cu (II) quercetin complexes suggests that Cu (I) and Cu (II) have significantly modified the chemical composition of the quercetin (Panhwar et al., 2010). The reason could be due to the decrease in oxidation potential of quercetin by Cu (I) and Cu (II) complexation thus, making it readily oxidized, more reactive and effective than free quercetin (Bukhari et al., 2008; Panhwar et al., 2010).

Conclusion

This study has reported the effects of redox states on syntheses, spectra characterizations, antioxidant activities of quercetin and its copper complexes. The complexes and quercetin exhibited different spectroscopic properties while complexes possessed better free radical scavenging ability than free quercetin with Cu (I)quercetin complex having the highest antioxidant activity.

COMPETING INTEREST

Authors have not declared any conflict of interest.

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